

Evaluation of Various Ionic Currents Generating Cardiac Action Potential by Patch-Clamp Method

心筋活動電位を形成する各種イオンチャネル電流のパッチクランプ法による評価

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Introduction

The hERG assay is now a familiar measure of predicting drug-induced QT prolongation in the preclinical stage. Recently effects on cardiac ion channels other than hERG is coming under a spot light, and many are suggesting assessing effects on those ion channels in an early stage. We investigated methods of assessing the currents through those ion channels by patch-clamp technique with stably expressing cell lines.

Materials and Methods

Cell Line

Cell Line	Supplier	Current
CHO-hCa _v 1.2/β ₂ /α ₂ δ ₁	ChanTest	I _{Ca}
HEK-hNa _v 1.5	SB Drug Discovery	I _{Na}
HEK-hK _v LQT1/minK	Cytocentrics	I _{Ks}
CHO-hK _v 2.1	B'SYS	I _{K1}
CHO-hK _v 1.5	Cytocentrics	I _{Kur}
CHO-hK _v 4.3	B'SYS	I _{to}

Patch Clamp

- Whole-cell patch-clamp method
- Application: single or cumulative
- Flow rate: 3-5 mL/min
- Pipette resistance: 1-5 MΩ

Solution

Extracellular solution

	I _{Na}	I _{Ca}	I _{Ks}	I _{K1}	I _{Kur}	I _{to}
NaCl	140	145	145		137	
KCl	4	4	4		4	
CaCl ₂	2	10	2		1.8	
MgCl ₂	1		1		1	
glucose	10	10	10		10	
HEPES	10	10	10		10	
pH	7.35	7.4	7.4		7.4	

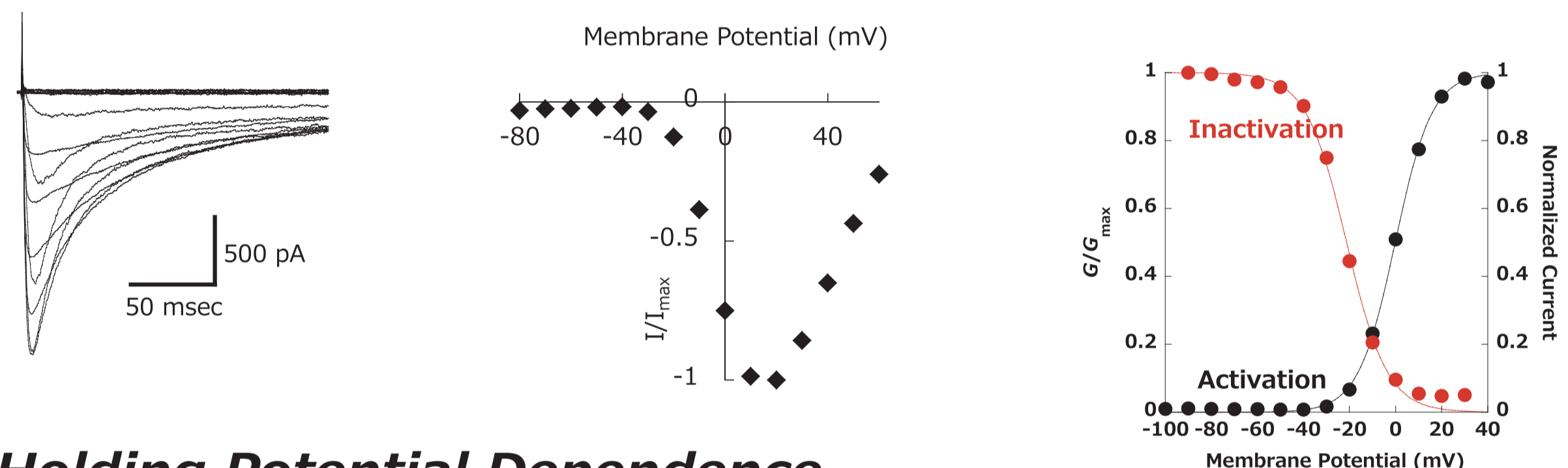
Intracellular solution

	I _{Na}	I _{Ca}	I _{Ks}	I _{K1}	I _{Kur}	I _{to}
KCl					130	
CsCl	50	112				
NaCl	10	2				
CsF	60	27				
MgCl ₂	2				1	
EGTA	20	8.2			5	
MgATP		4			5	
HEPES	10	10			10	
pH	7.25	7.2			7.2	

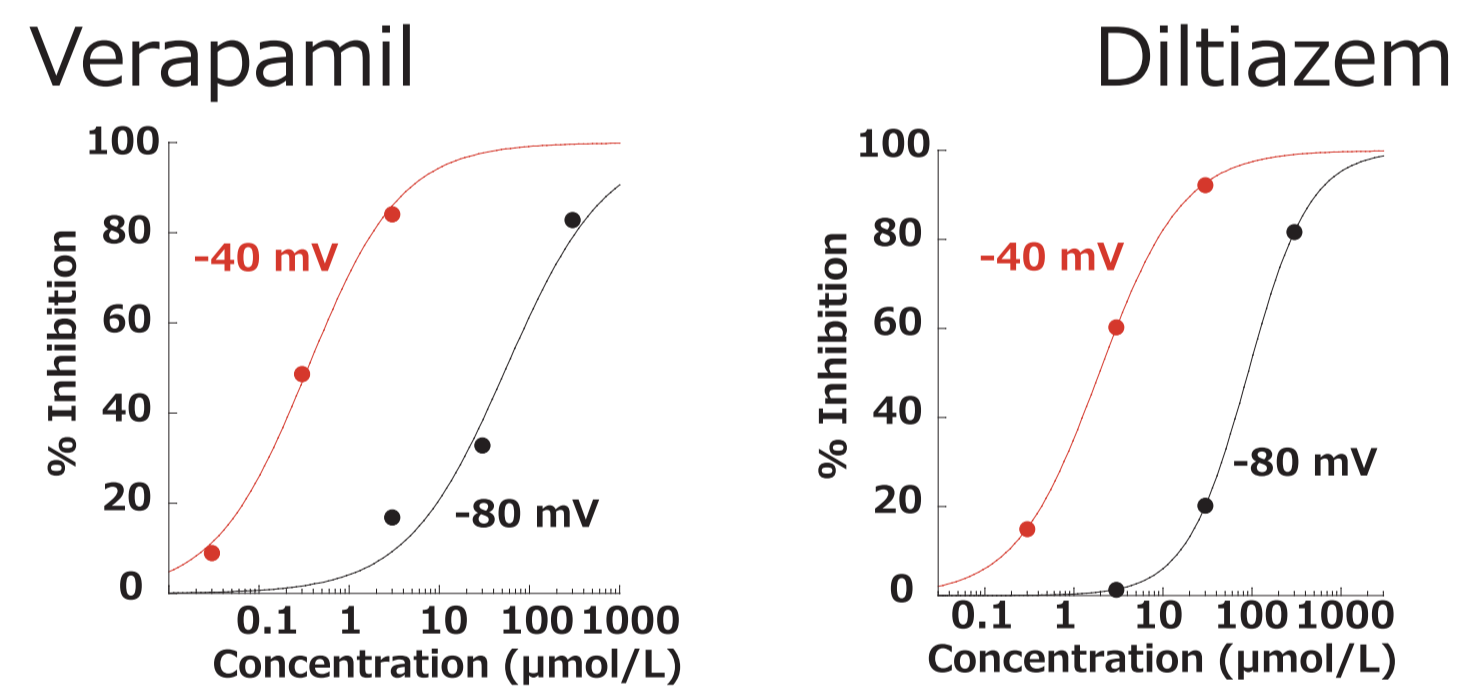
Results

Ca_v1.2

Electrophysiological Property



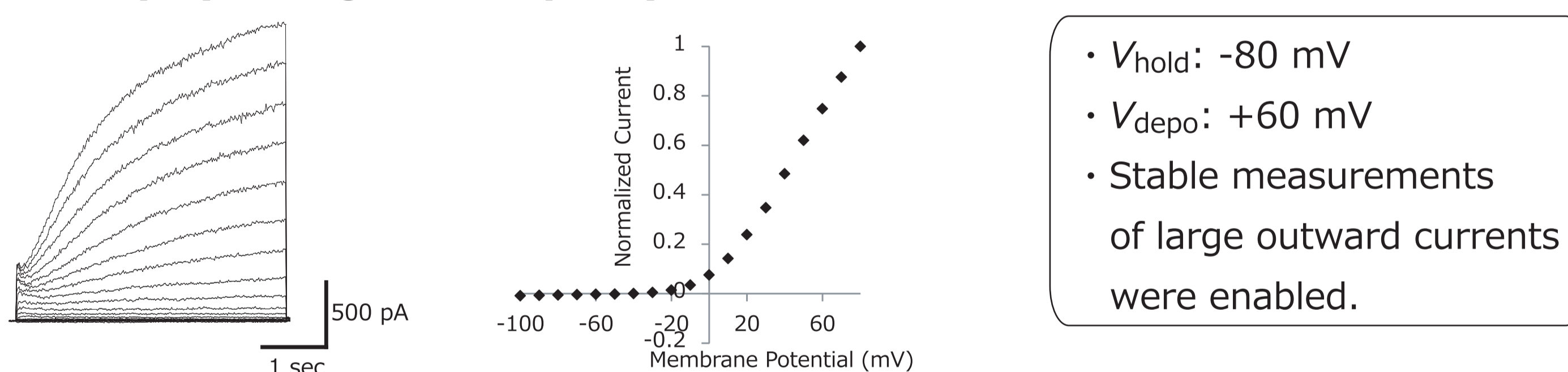
Holding Potential Dependence



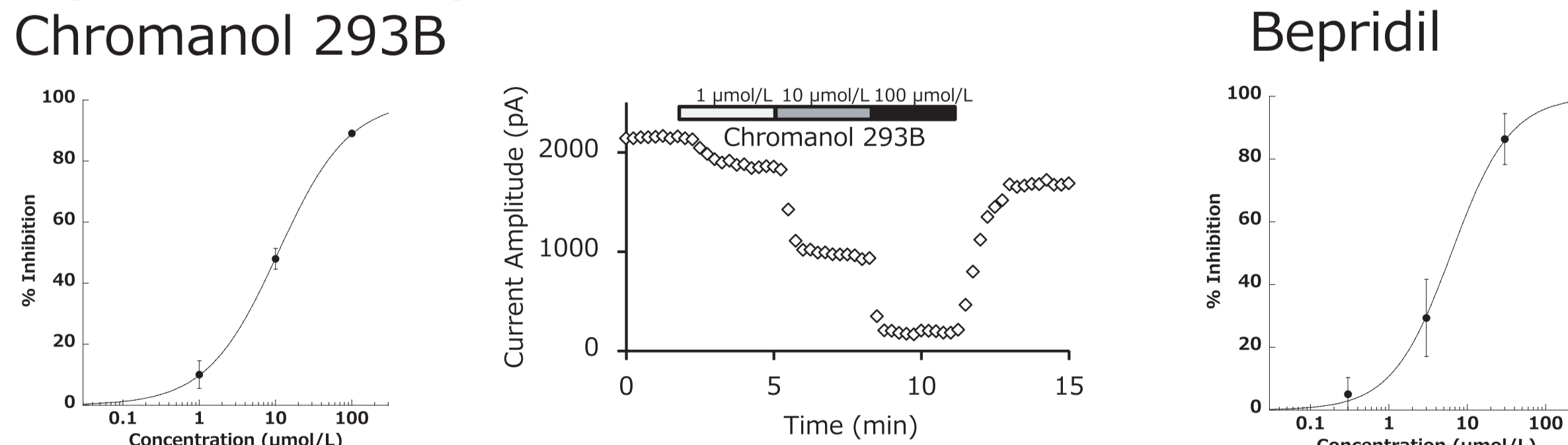
- V_{hold}: -80 mV or -40 mV
- V_{depo}: 0 mV
- Inhibitory activities of compounds depend on holding potentials.

K_vLQT1/minK

Electrophysiological Property



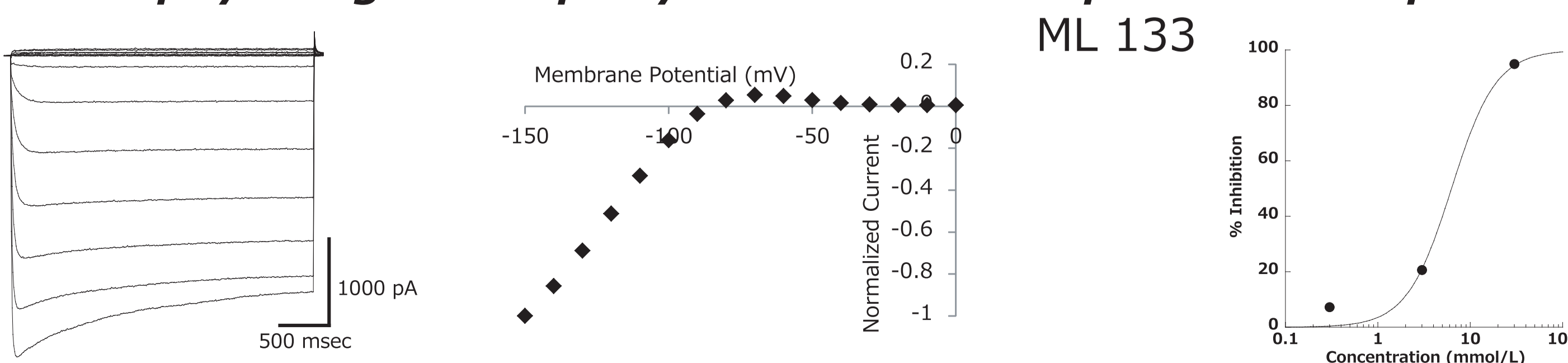
Response to Compounds



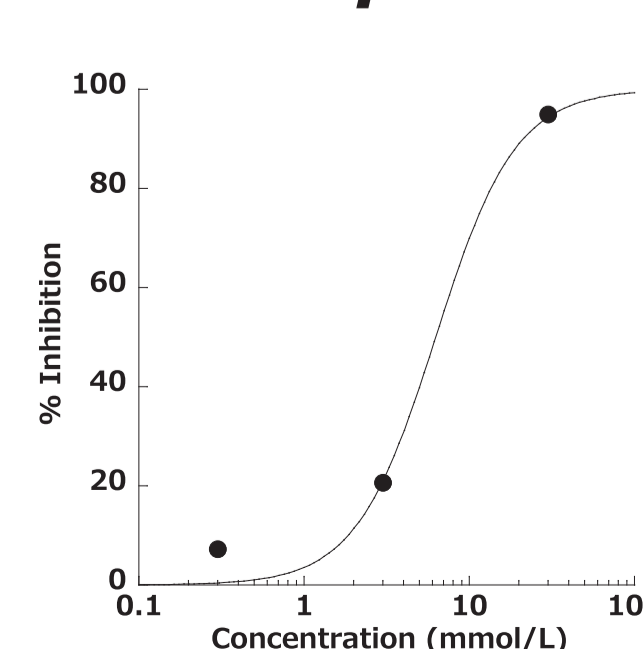
- V_{hold}: -80 mV
- V_{depo}: +60 mV
- Stable measurements of large outward currents were enabled.

K_{ir}2.1

Electrophysiological Property



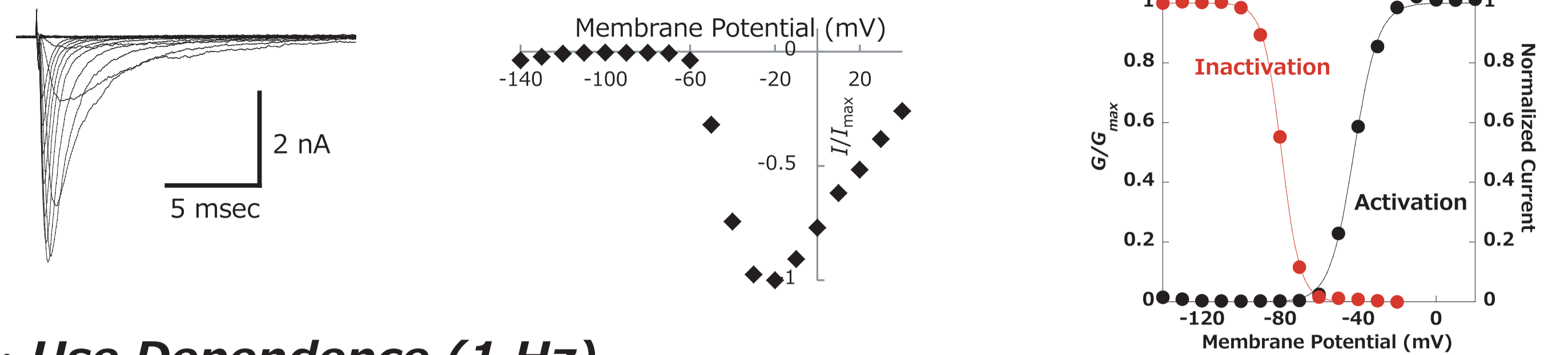
Response to Compounds



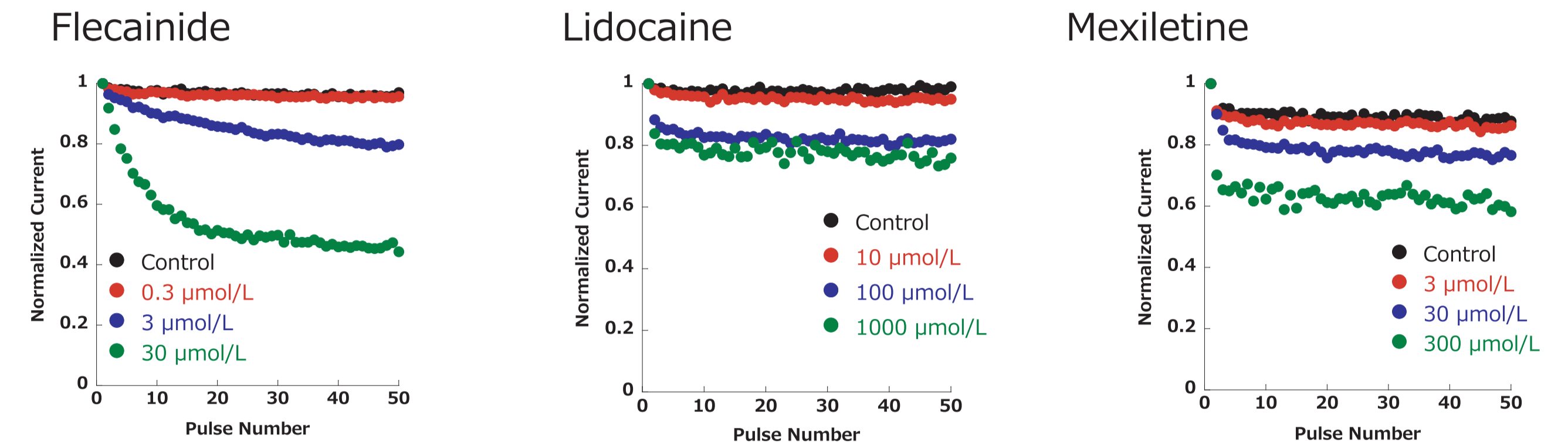
- V_{hold}: -20 mV
- V_{hypo}: -120 mV
- Notable inward-rectifying current was identified.

Na_v1.5

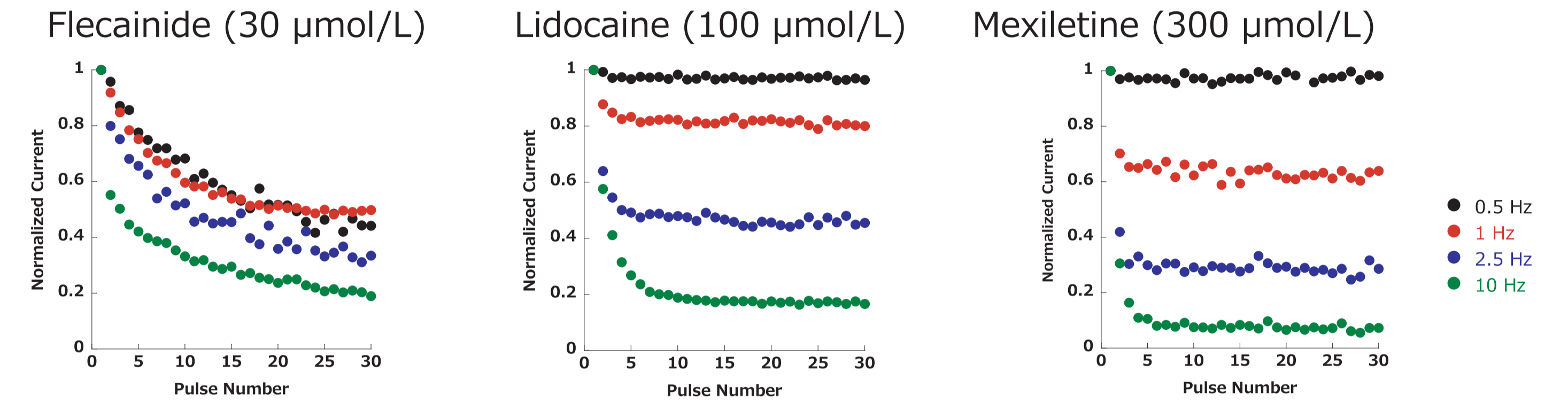
Electrophysiological Property



Use Dependence (1 Hz)



Frequency Dependence



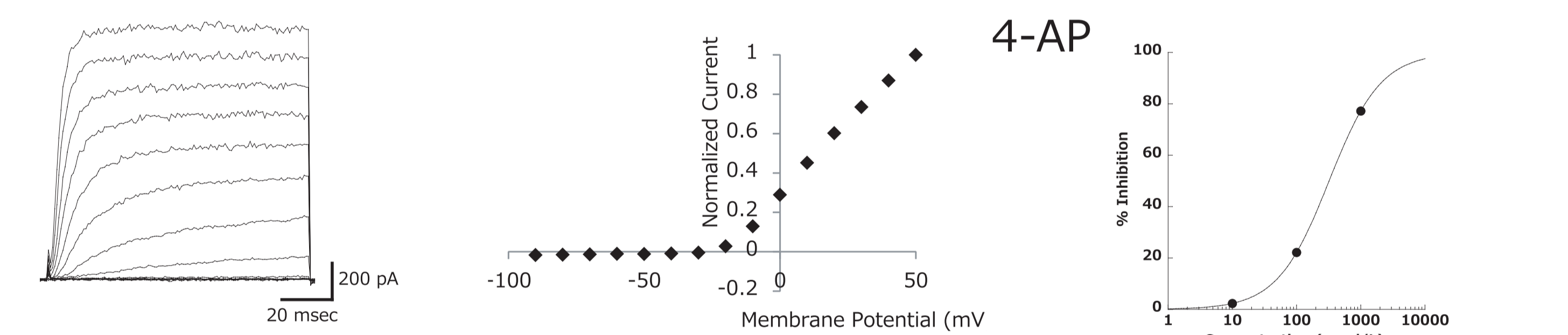
Results from various protocols (comparison of IC₅₀s; μmol/L)

Protocol	Flecainide	Lidocaine	Mexiletine
Tonic Block	> 30	274.4	87.0
UD 0.5 Hz	50 _{th} /1 _{st}	19.6	> 1000
	50 _{th} _{cpd} /50 _{th} _{ctrl}	5.6	466.8
UD 1 Hz	50 _{th} /1 _{st}	23.6	> 1000
	50 _{th} _{cpd} /50 _{th} _{ctrl}	6.2	141.6
UD 2.5 Hz	50 _{th} /1 _{st}	11.7	> 1000
	50 _{th} _{cpd} /50 _{th} _{ctrl}	2.7	31.3
UD 10 Hz	50 _{th} /1 _{st}	8.7	25.6
	50 _{th} _{cpd} /50 _{th} _{ctrl}	4.3	21.5
Inactivation	> 30	18.2	15.0

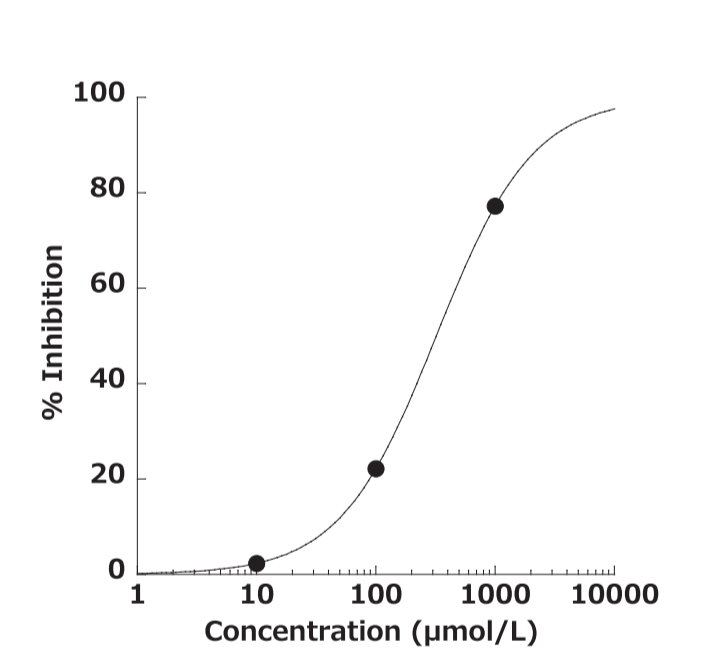
- V_{hold}: -100 mV
- V_{depo}: -20 mV
- Inhibitory activities of compounds depend on setting protocols.

K_v1.5

Electrophysiological Property



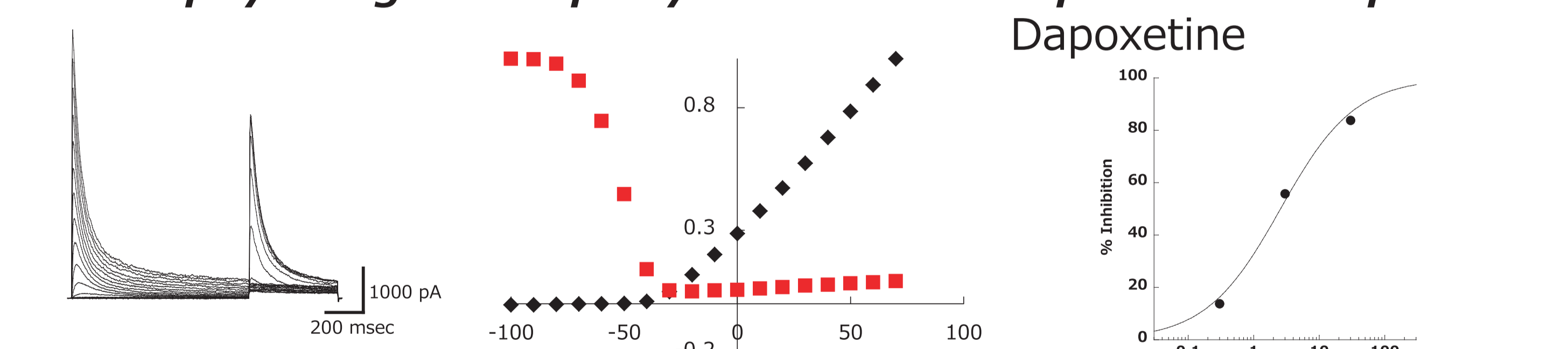
Response to Compounds



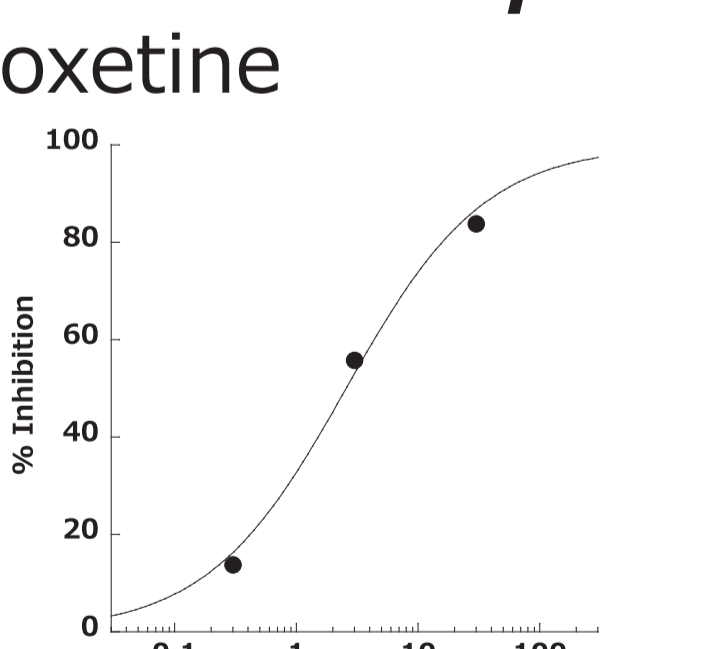
- V_{hold}: -80 mV
- V_{depo}: +20 mV
- Stable measurements of large outward currents were enabled.

K_v4.3

Electrophysiological Property



Response to Compounds



- V_{hold}: -80 mV
- V_{depo}: +20 mV
- Stable measurements of transient outward currents were enabled.

Conclusion

Now we are able to perform stable measurements of cardiac ion-channel currents. Protocols and experimental conditions must be optimized depending on purposes, since the degree of inhibitory activity depends on experimental setting. For evaluating effects of compounds on these cardiac ion channels, standardization of experimental conditions appropriate to the evaluation purpose (safety pharmacology, target assay, etc), based on knowledge of both electrophysiological and pharmacological properties of each ion channel, would be necessary.